

## Echinacea purpurea Aerial Parts

### DEFINITION

*Echinacea purpurea* Aerial Parts consists of the aerial parts of *Echinacea purpurea* (L.) Moench (Fam. Asteraceae). It is harvested during the flowering stage. It contains NLT 1.0% of the sum of caftaric acid (C<sub>13</sub>H<sub>12</sub>O<sub>9</sub>) and chicoric acid (C<sub>22</sub>H<sub>18</sub>O<sub>12</sub>), and NLT 0.01% of dodecatetraenoic acid isobutylamides (C<sub>16</sub>H<sub>25</sub>NO), calculated on the dried basis.

### IDENTIFICATION

#### A. THIN-LAYER CHROMATOGRAPHY

**Presence of chicoric acid and absence of echinacoside**

**Standard solution A:** 0.2 mg/mL of USP Echinacoside RS in methanol

**Standard solution B:** 0.2 mg/mL of USP Caftaric Acid RS, 0.1 mg of USP Chlorogenic Acid RS, and 0.2 mg/mL of USP Chicoric Acid RS in methanol

**Standard solution C:** 20 mg/mL of USP Powdered *Echinacea purpurea* Extract RS in methanol. Shake to disperse, sonicate for 5 min, and centrifuge. Use the supernatant.

**Sample solution:** Transfer 1 g of finely pulverized *Echinacea purpurea* Aerial Parts to a centrifuge tube, add 10 mL of methanol, mix well, and sonicate for 10 min. Centrifuge, and use the supernatant.

#### Chromatographic system

(See *Chromatography* <621>, *System Suitability*.)

**Adsorbent:** Chromatographic silica gel mixture with an average particle size of 5 µm (HPTLC plates)

**Application volume:** 5 µL *Standard solution C* and *Sample solution*, and 2 µL *Standard solution A* and *Standard solution B* as 8-mm bands

**Relative humidity:** Condition the plate to a relative humidity of about 33% using a suitable device.

**Developing solvent system:** A mixture of ethyl acetate, methylethyl ketone, water, and formic acid (5:3:1:1)

**Developing distance:** 6 cm

**Derivatization reagent:** 5 mg/mL of 2-aminoethyl diphenylborinate in ethyl acetate

#### Analysis

**Samples:** *Standard solution A*, *Standard solution B*, *Standard solution C*, and *Sample solution*

Apply the *Samples* as bands to a suitable thin-layer chromatographic plate, and dry in air. Develop the chromatograms in a saturated chamber. Remove the plate from the chamber, heat at 100° for 5 min, derivatize the plate while still warm with *Derivatization reagent*, dry in air, and examine under UV light at 366 nm.

**System suitability:** *Standard solution A* shows one major blue band in the lower third section of the chromatogram due to echinacoside. *Standard solution B* shows two major blue bands at about the middle of the chromatogram due to caftaric acid (higher *R<sub>F</sub>*) and chlorogenic acid (lower *R<sub>F</sub>*) that are clearly separated, and a blue band for chicoric acid in the upper third section of the chromatogram.

**Acceptance criteria:** The most prominent band in the *Sample solution* chromatogram is a blue band in the upper third section of the chromatogram at an *R<sub>F</sub>* corresponding to the chicoric acid band in the chromatogram of *Standard solution B* and *Standard solution C*. The second most prominent band in the *Sample solution* chromatogram is a blue band at about the middle of the chromatogram due to caftaric acid, corresponding to a band in the chromatogram of *Standard solution C*. The *Sample solution* chromatogram does not exhibit a band at the *R<sub>F</sub>* of echinacoside in *Standard solution A* (difference from *Echinacea pallida* and *Echinacea angustifolia*). The *Sample solution* chromatogram exhibits minor blue bands

corresponding to similar bands in the chromatogram of *Standard solution C*. One of these bands is due to chlorogenic acid at an *R<sub>F</sub>* corresponding to chlorogenic acid in *Standard solution B*. The *Sample solution* chromatogram exhibits a red band due to chlorophyll close to the solvent front.

- B.** The retention time of the major peak in the *Sample solution* corresponds to that of the chicoric acid peak in *Standard solution A*, and the second most prominent peak corresponds to that of the caftaric acid peak in *Standard solution B*. The *Sample solution* chromatogram shows no peak or a very minor peak at the retention time corresponding to the echinacoside peak in the *Standard solution C* chromatogram, all peaks as obtained in the test for *Content of Chicoric Acid and Caftaric Acid*.
- C.** The retention times for the relevant peaks of the *Sample solution*, mainly due to dodecatetraenoic isobutyl amides, correspond to those of *Standard solution A*, as obtained in the test for *Content of Dodecatetraenoic Isobutylamides*.

### COMPOSITION

#### CONTENT OF CHICORIC ACID AND CAFTARIC ACID

**Solution A:** Phosphoric acid (0.1 in 100) in water

**Solution B:** Acetonitrile

**Mobile phase:** See *Table 1*.

**Table 1**

Time (min)	Solution A (%)	Solution B (%)
0	90	10
13	78	22
14	60	40
17.5	60	40
18	90	10
30	90	10

**Solvent:** Alcohol and water (7:3)

**Standard solution A:** 30 µg/mL of USP Chicoric Acid RS in *Solvent*

**Standard solution B:** 20 µg/mL of USP Caftaric Acid RS in *Solvent*

**Standard solution C:** 20 µg/mL of USP Echinacoside RS in *Solvent*

**Sample solution:** Transfer about 125 mg, accurately weighed, of finely powdered *Echinacea purpurea* Aerial Parts (capable of passing through a 40-mesh sieve) to a round-bottom flask equipped with a condenser. Add 25.0 mL of *Solvent*, and heat under reflux while shaking by mechanical means for 15 min. Centrifuge, or pass through a membrane filter of 0.45-µm or finer pore size.

#### Chromatographic system

(See *Chromatography* <621>, *System Suitability*.)

**Mode:** LC

**Detector:** UV 330 nm

**Column:** 4.6-mm × 25-cm; 5-µm packing L1

**Column temperature:** 35°

**Flow rate:** 1.5 mL/min

**Injection size:** 5 µL

#### System suitability

**Samples:** *Standard solution A*

#### Suitability requirements

**Relative standard deviation:** NMT 2.0% for the chicoric acid peak in *Standard solution A*

#### Analysis

**Samples:** *Standard solution A*, *Standard solution B*, *Standard solution C*, and *Sample solution*

Separately calculate the percentages of caftaric acid ( $C_{13}H_{12}O_9$ ) and chicoric acid ( $C_{22}H_{18}O_{12}$ ) in the portion of *Echinacea purpurea* Aerial Parts taken:

$$\text{Result} = (r_U/r_S) \times C_S \times (V/W) \times 100$$

- $r_U$  = peak area of the relevant analyte from the *Sample solution*  
 $r_S$  = peak area of the relevant analyte from the corresponding *Standard solution*  
 $C_S$  = concentration of the relevant analyte in the corresponding *Standard solution* (mg/mL)  
 $V$  = final volume of the *Sample solution* (mL)  
 $W$  = weight of *Echinacea purpurea* Aerial Parts taken to prepare the *Sample solution* (mg)

Calculate the percentage of the sum of chicoric acid and caftaric acid in the portion of *Echinacea purpurea* Aerial Parts taken by adding the individual percentages calculated.

**Acceptance criteria:** NLT 1.0% on the dried basis

• **CONTENT OF DODECATETRAENOIC ACID ISOBUTYLAMIDES**

**Mobile phase:** Acetonitrile and water (55:45)

**Standard solution A:** 5 mg/mL of USP Powdered *Echinacea purpurea* Extract RS in methanol. Dissolve using sonication and shaking for 10 min. After dilution, pass through a membrane filter of 0.45- $\mu$ m or finer pore size.

**Standard solution B:** 10  $\mu$ g/mL of USP 2*E*,4*E*-Hexadienoic Acid Isobutylamide RS in methanol

**Sample solution:** Transfer about 2.5 g of finely powdered *Echinacea purpurea* Aerial Parts (capable of passing through a 40-mesh sieve), accurately weighed, into a round-bottom flask. Add 80 mL of methanol, and reflux for 30 min. Cool to room temperature, and filter into a 100-mL volumetric flask, using small portions of methanol to rinse the flask and the filter. Dilute with methanol to volume. Pass through a membrane filter of 0.45- $\mu$ m or finer pore size.

**Chromatographic system**

(See *Chromatography* (621), *System Suitability*.)

**Mode:** LC

**Detector:** UV 254 nm

**Column:** 4.6-mm  $\times$  25-cm; 5- $\mu$ m packing L1

**Column temperature:** 30°

**Flow rate:** 1.5 mL/min

**Injection size:** 25  $\mu$ L

**System suitability**

**Samples:** *Standard solution A* and *Standard solution B*

**Suitability requirements**

**Chromatogram similarity:** The chromatogram from *Standard solution A* is similar to the Reference Chromatogram for alkaloids provided with USP Powdered *Echinacea purpurea* Extract RS.

**Resolution:** NLT 1.0 between dodecatetraenoic acid isobutylamide peaks, *Standard solution A*

**Tailing factor:** NMT 2.0 for 2*E*,4*E*-hexadienoic acid isobutylamide, *Standard solution B*

**Relative standard deviation:** NMT 2.5% for the 2*E*, 4*E*-hexadienoic acid isobutylamide peak in repeated injections, *Standard solution B*

**Analysis**

**Samples:** *Standard solution A*, *Standard solution B*, and *Sample solution*

Identify the peaks of the two isomers of dodecatetraenoic acid isobutylamides in the chromatogram from the *Sample solution* by comparison with the chromatogram from *Standard solution A*. Measure the areas for the relevant peaks.

Calculate the percentage of dodecatetraenoic acid isobutylamides in the portion of *Echinacea purpurea* Aerial Parts taken:

$$\text{Result} = (r_U/r_S) \times C_S \times (V/W) \times F \times 100$$

- $r_U$  = sum of the peak areas of the relevant analytes from the *Sample solution*  
 $r_S$  = peak area of 2*E*,4*E*-hexadienoic acid isobutylamide from *Standard solution B*  
 $C_S$  = concentration of USP 2*E*,4*E*-Hexadienoic Acid Isobutylamide RS in *Standard solution B* (mg/mL)  
 $V$  = final volume of the *Sample solution* (mL)  
 $W$  = weight of *Echinacea purpurea* Aerial Parts taken to prepare the *Sample solution* (mg)  
 $F$  = response factor to convert 2*E*,4*E*-hexadienoic acid isobutylamide into dodecatetraenoic acid isobutylamides, 1.353

**Acceptance criteria:** NLT 0.01% of dodecatetraenoic acid isobutylamides on the dried basis

**CONTAMINANTS**

• **ELEMENTAL IMPURITIES—PROCEDURES** (233)

**Acceptance criteria**

**Arsenic:** NMT 1.0  $\mu$ g/g

**Cadmium:** NMT 0.5  $\mu$ g/g

**Lead:** NMT 5.0  $\mu$ g/g

**Mercury:** NMT 1.0  $\mu$ g/g

**Change to read:**

- **ARTICLES OF BOTANICAL ORIGIN, Pesticide Residue Analysis** (561)  $\Delta$  (CN 1-May-2019): Meet the requirements
- **MICROBIAL ENUMERATION TESTS** (2021): The total aerobic microbial count does not exceed  $10^5$  cfu/g, the total combined molds and yeasts count does not exceed  $10^3$  cfu/g, and the enterobacterial count does not exceed  $10^3$  cfu/g.
- **ABSENCE OF SPECIFIED MICROORGANISMS** (2022): It meets the requirements of the tests for absence of *Salmonella* species and *Escherichia coli*.

**SPECIFIC TESTS**

• **BOTANIC CHARACTERISTICS**

**Macroscopic:** The herb is an erect, coarse, rough-hairy perennial, usually up to 90 cm tall, rarely up to 180 cm. The leaves are alternate and simple; the lowermost leaves are slender, long, and petioled, ovate to broadly lanceolate, mostly penta-nerved, acute or acuminate at the apex, abruptly narrowed or rarely cordate at the base, usually sharply dentate, and 7–20 cm long and 2.5–7.5 cm wide; the petioles are mostly winged at the summit. The upper leaves are narrower, often almost entirely sessile, lanceolate or ovate lanceolate, and usually with 3 veins. The flower heads are radiate, up to 15 cm across, solitary or few, and long-peduncled, with 12–20 rays, purple, crimson, or rarely pale; the bristle disks are often orange, 3.5–7.5 cm long; the involucre is depressed-hemispheric; the bracts are lanceolate, spreading or appressed, imbricated in 2–4 series, and hairy on the outer surface with ciliate margins; the receptacle is conical, the scales of the receptacle stiff, spinescent, and conspicuously longer than the disc flowers; the chaff is carinate and cuspidate; the achenes are 3–4 mm in length, tetrasided, obypyramidal, and thick; the pappus has a short, dentate crown.

**Microscopic**

**Leaf:** The leaf has a thickness of 200–350  $\mu\text{m}$ , with an epidermis 9–13  $\mu\text{m}$  thick, largely without chloroplasts; the stomata are 28–35  $\mu\text{m}$ , abundant on the ventral surface and fewer on the dorsal surface; the mesophyll is clearly divided into palisade parenchyma and sponge parenchyma. The palisade parenchyma is one layer thick, with elongated cells 50–65  $\mu\text{m}$  in length, oriented at right angles to the leaf surface, containing numerous chloroplasts. The sponge parenchyma is 150–250  $\mu\text{m}$  thick, with cells of irregular shape, and has multiple cell layers, few chloroplasts, and large intercellular spaces. The phloem bundles of the lateral veins within the sponge parenchyma are bound by a one-layer sheath of small parenchymous cells, with vascular elements of the midrib surrounded by large-celled parenchyma. The uniseriate trichomes are few in the ventral surface, numerous on the dorsal surface, typically tricelled, occasionally tetra- or pentacelled, 250–500  $\mu\text{m}$  in length, each arising from an epidermal cell; the epidermal cell walls appear with moderate thickening; the vessels are various, scalariform, with variable reticulated width.

**Petiole:** The parenchyma appear without chloroplasts, in several layers adjacent to a layer of collenchyma; 5–7 phloem bundles of small- to medium-sized vessels are weakly lignified and embedded in the parenchyma in the form of an arc; the wing ribs of the upper surface of the slightly hollowed petiole are marginal.

**Inflorescence:** The epidermal cells of the ray florets are square, 50  $\mu\text{m}$ , with a transparent, beaded cell wall; various elements of the *Asteraceous* exhibit inflorescence; numerous multicellular jointed trichomes of the involucre bracts are 500–800  $\mu\text{m}$  in length; tangential sections of

the paleae with numerous fiber bundles are 10–15  $\mu\text{m}$  in diameter and 100–150  $\mu\text{m}$  in length; cell walls are thin. The epidermis of ray florets is reddish to violet; the epidermal cells from the end of the corolla form rounded papillae; a stigma of papillary cells is present; *Asteraceous* pollen grains are 20–30  $\mu\text{m}$  and spherical with a warty exine.

Calcium oxalate is negative; crystals of inulin and starch granules are rare.

- **ARTICLES OF BOTANICAL ORIGIN**, *Foreign Organic Matter* (561): NMT 3.0%

- **LOSS ON DRYING** (731)

**Sample:** 1 g of the powdered plant material

**Analysis:** Dry the *Sample*.

**Acceptance criteria:** NMT 12%

- **ARTICLES OF BOTANICAL ORIGIN**, *Total Ash* (561): NMT 10.0%, determined on 3 g

- **ARTICLES OF BOTANICAL ORIGIN**, *Acid-Insoluble Ash* (561): NMT 2.5%

**ADDITIONAL REQUIREMENTS**

- **PACKAGING AND STORAGE:** Store in tight, light-resistant containers at controlled room temperature.
- **LABELING:** The label states the Latin binomial and, following the official name, the parts of the plant contained in the article.
- **USP REFERENCE STANDARDS** (11)
  - USP Chlorogenic Acid RS
  - USP Caftaric Acid RS
  - USP Chicoric Acid RS
  - USP Echinacoside RS
  - USP 2*E*,4*E*-Hexadienoic Acid Isobutylamide RS
  - USP Powdered *Echinacea purpurea* Extract RS